

Enzyme REACH Consortium Guidance: Application of read-across for enzyme substances

1. Introduction

Enzymes are protein with catalytic activities and belong to Unknown or Variable Composition, Complex Reaction Products, or Biological Materials (UVCB). Enzyme substances are enzyme concentrate, dry matter obtained after the manufacturing process and they are identified based on catalytic activities defined in International Union of Biochemistry and Molecular Biology (IUBMB). Enzyme concentrates with the same IUBMB number can be regarded as the same substance, despite using different production organism, provided that the hazardous properties do not differ significantly and warrant the same classification. (ECHA's guidance for identification and naming of substances under REACH and CLP, Section 4.3.2.3 and Section 5). Enzyme REACH Consortium (ERC) further developed industrial guidance how to document the same classification within the same IUBMB (ERC Safety evaluation of technical enzyme products with regards to the REACH legislation [1]).

Enzymes registered under REACH regulation must fulfil information requirements that depend on the tonnage for which the substance is registered. Information requirements can be fulfilled by conducting testing on the substance itself. Other options include using existing data (generated for purposes other than REACH), weight of evidence approach, Qualitative or Quantitative structure-activity relationship (QSAR) modelling or grouping of substances and read-across approach. In some cases, it is also possible to waive the information requirement if the testing is not technically or scientifically possible (see ERC document on Data waiving argumentation for technical enzymes).

1.1. Selecting read-across approach

For enzymes, read-across can be utilised for physico-chemical, toxicological and environmental toxicity endpoints. Depending on the endpoint and the data availability, either **analogue** or **category approach** can be used for read-across.

In the **analogue approach** test data from a structural analogue (*source* substance) is used to predict the properties of the registered substance (*target* substance) i.e., one-to-one read-across. For this approach it is recommended to use an enzyme in the same IUBMB sub-sub-class, or at least sub-class, as a source substance (e.g., source substance alpha-amylase IUBMB 3.2.1.1 for target substance cellulase IUBMB 3.2.1.4) to support the argument of structural similarity.

In the **category approach** read-across is applied between the registered substance (*target* substance) and a group of structurally similar substances, i.e., many-to-one read-across. Substances in the group should be grouped together on defined structural similarities and/or differences. With the category approach it is proposed that the properties of the substances are either similar or they follow a pattern that can be applied also to the target substance. Furthermore, it is recommended to build the category from enzymes that at least belong to same IUBMB class to support structural similarity.

Whether an analogue or category approach is selected for the read-across, the justification for the selection needs to be documented in a read-across assessment report. Templates for reporting are made

available by ECHA in their Guidance on information requirements and chemical safety assessment Chapter R.6: QSARs and grouping of chemicals section R.6.2.6 [2].

For further general information on read-across approach, see ECHA guidance documents Read-Across Assessment Framework (RAAF) [3] and RAAF: Considerations on multi-constituent substances and UVCBs [4] (March 2017) and Advice on using read-across for UVCB substances [5] (May 2022).

2. Physico-chemical properties

Most of the physico-chemical properties are waived for enzymes (see document Data waiving argumentation for technical enzymes). However, for a few endpoints read-across is a valid option to fulfil information requirements. With regards to physico-chemical properties, all enzymes in different IUBMB classes can be grouped together, as the physico-chemical properties of enzyme concentrate, dry matter are independent to the catalytic activity of the enzyme but based on the fact that all enzymes are proteins. The dry concentrate form of all enzyme substances is shown to act similarly in the tests required for REACH.

2.1. Density

Density data have been generated for various protein families. The data published in the cited book comes from 20 different protein families and they all have densities in the range from 1.32 g/mL to 1.42 g/mL. The density for the enzyme has not been determined but since it is a protein it is expected, with very high reliability that it has density in this range.

Thomas E. Creighton. (1993). Proteins: structures and molecular properties. W.H.Freeman and Company. ISBN 0-7167-2317-4

2.2. Vapour pressure

Enzymes are protein and vapor pressures of 4 different enzymes (alpha-amylase, cellulase, glucoamylase and subtilisin) were measured with freeze-dried test samples. The vapor pressures of the 4 enzymes were indeed very similar from 0.00195 to 0.00689 Pa. Other enzymes are also protein, thereby vapour pressure is expected to be very similar to the 4 enzymes, therefore these data are used for read-across.

2.3. Partition coefficient n-octanol/water

Because all enzymes are built up of the combination of the same 20 common amino acids, the physical and chemical characteristics are very similar for different enzymes, and hence, read-across from other enzymes should be fully applicable. Enzymes have been analysed and the LogPow from literature studies was found to be between -3.1 to -2.95. In addition, the octanol-water partition coefficient (logPow) for glucoamylase was measured <= -1.3 at 20°C.

Due to the similar nature of enzymes, this value can also be extrapolated to other enzymes.

References:

- Basketter D, Berg N, Broekhuizen C, Fieldsend M, Kirkwood S, Kluin C, Mathieu S, Rodriguez C (2012a). Enzymes in Cleaning Products: An Overview of Toxicological Properties and Risk Assessment/Management. *Regul. Toxicol. Pharmacol.*, 64(1):117-123.

2.4. Flammability

Read-across data on substance cellulase (IUBMB 3.2.1.4) has been made available in 2020 for the endpoint vapour pressure for all ERC members who act as lead registrants. As there is only one source substance data, an analogue approach is suggested for this endpoint.

Cellulase was freeze-dried and tested for flammability. The test showed that cellulase is not flammable.

Most of the enzymes are produced by fermentation, typically in solution. After the fermentation, enzymes are recovered as enzyme concentrate in a liquid form and further formulated as liquid products or granules. The substance in contact with water is not considered to be highly flammable according to Regulation (EC) No 440/2008. Based on this data, other enzymes are also expected to be Non-flammable (Not Classified).

Enzymes are globular proteins produced by fermentation i.e. typically in solution. After the fermentation, enzymes are recovered as enzyme concentrate in a liquid form and further formulated as liquid products or granules. Decades of experience in production, handling and use of enzymes show that the substance does not react with water (e.g. the substance is manufactured with water or/and washed with water). The substance is soluble and stable in water. Therefore, the substance in contact with water is not considered to be highly flammable according to Regulation (EC) No 440/2008. Under REACH, enzymes are defined as enzyme concentrate, dry matter [6]. As dry matter, proteins in general are not considered to be highly flammable according to Regulation (EC) No 440/2008. This is supported by the chemical structure of the proteins. Proteins contain reactive groups such as hydroxyl, carboxylic acid, amines, thiols groups etc. During combustion, carboxylic acid groups may go through decarboxylation and hydroxyl groups may be released as water vapor [7]. The carboxylic acid may also promote char formation [7, 8]. Proteins also contain nitrogen and sulfur which form a disulfide bond helping in contributing to its non-inherent flammability [7, 8]. These properties have in fact led to the use of proteins as flame retardants [7, 8]. Additionally, several proteins have already been tested using recognized method under REACH and CLP (e.g. UN Test N.1) and found not to be highly flammable according to Regulation (EC) No 440/2008, e.g. Protein hydrolyzates, rice bran (EC number: 305-224-5), Protein hydrolyzates, animal (EC number: 309-203-1), Insulin DesB30 (EC number: 944-550-8).

3. Toxicological properties

In general, enzymes exhibit the same toxicological properties and besides the fact that they are respiratory sensitizers are of low toxicity, which is confirmed by toxicity studies performed in the industry and published safety evaluations for a variety of enzymes [9-53].

Read across for toxicological endpoints can be applied for enzyme substances that belong to same IUBMB sub-sub-class or sub-class, provided that the safety of the other constituents has been established (safe-strain lineage) and that the toxic effect with regards to the selected endpoint can be considered comparable. However, read-across between enzymes of different IUBMB class should only be considered and performed when there are no studies available for the specific enzyme. In general, read-across between enzymes of different IUBMB numbers can be performed for the majority of enzyme substances, except for proteases due to the intrinsic properties of proteases to catalyse protein degradation. In order to perform read-across between enzymes of different IUBMB classification for health hazard identifications, except genotoxicity, enzyme substances can be divided into two groups, proteases and non-proteases. For genotoxicity, all enzymes can be grouped together since the overall conclusion is that enzymes are not mutagenic or clastogenic.

For acute skin and eye irritation as well as genotoxicity, qualitative read-across can be applied since there is typically no dose response relationship. Qualitative and quantitative read-across can be applied for acute oral and repeated dose oral toxicity.

4. Ecotoxicological properties

In general, enzymes exhibit the same ecotoxicological properties which are confirmed by ecotoxicity studies performed in the industry.

In general, the same rules for read-across for health hazards identification described previously can be applied for environmental hazard identification. However, to perform read-across between enzymes of different IUBMB classes for environmental hazards, enzyme substances should be divided into three groups, proteases, oxidoreductases and all other enzymes. With regards to biodegradability all enzymes can be grouped together.

Qualitative read-across will be applied for both aquatic toxicity and ready biodegradability since the PNEC (predicted no effect concentration) values for the majority of enzymes, except for proteases and oxidoreductases, is considered the highest dose tested for a given end point. As far as biodegradability is concerned all enzymes are considered readily biodegradable according to the current OECD guidelines indicating a simple yes or no answer without a quantitative relationship. Aquatic toxicity tests should be performed according to current OECD guidelines and under consideration that enzymes are readily biodegradable i.e., semi-static system when possible.

References:

- (1) ERC Safety evaluation of technical enzyme products with regard to REACH - SEPT 2021 .doc.pdf (enzymes-reach.org) last accessed 31 October 2024
- (2) https://echa.europa.eu/documents/10162/17224/information_requirements_r6_en.pdf/77f49f81-b76d-40ab-8513-4f3a533b6ac9?t=1322594777272 last accessed 31 October 2024
- (3) [614e5d61-891d-4154-8a47-87efebd1851a \(europa.eu\) last accessed 31 October 2024](https://echa.europa.eu/documents/10162/17224/information_requirements_r6_en.pdf/77f49f81-b76d-40ab-8513-4f3a533b6ac9?t=1322594777272)
- (4) [¹ 3f79684d-07a5-e439-16c3-d2c8da96a316 \(europa.eu\) last accessed 31 October 2024](https://echa.europa.eu/documents/10162/17224/information_requirements_r6_en.pdf/77f49f81-b76d-40ab-8513-4f3a533b6ac9?t=1322594777272)
- (5) [ac1f64a6-9ee5-441e-cf1c-92914b843b4e \(europa.eu\) last accessed 31 October 2024](https://echa.europa.eu/documents/10162/17224/information_requirements_r6_en.pdf/77f49f81-b76d-40ab-8513-4f3a533b6ac9?t=1322594777272)
- (6) Guidance for identification and naming of substances under REACH and CLP, Version 2.1.
- (7) <https://onlinelibrary.wiley.com/doi/full/10.1002/fam.2386>
- (8) <https://patents.google.com/patent/WO2000029662A1/en>
- (9) * Food and Agriculture Organization of the United Nations, * World Health Organization. Evaluation of certain food additives. World Health Organ Tech Rep Ser 2005; 928:1-156.
- (10) Ahmad SK, Brinch DS, Friis EP, Pedersen PB. Toxicological studies on Lactose Oxidase from *Microdochium nivale* expressed in *Fusarium venenatum*. *Regul Toxicol Pharmacol* 2004; 39(3):256-270.
- (11) Amalfitano A, Bengur AR, Morse RP et al. Recombinant human acid alpha-glucosidase enzyme therapy for infantile glycogen storage disease type II: Results of a phase I/II clinical trial. *Genet Med* 2001; 3(2):132-138.
- (12) Andersen JR, Diderichsen BK, Hjortkjaer RK et al. DETERMINING THE SAFETY OF MALTOGENIC AMYLASE PRODUCED BY RECOMBINANT DNA TECHNOLOGY. *J Food Prot* 1987; 50(6):521-526.
- (13) Ankel EG, Zirneski J, Ring BJ, Holcenberg JS. Effect of asparaginase on cell membranes of sensitive and resistant mouse lymphoma cells. *In Vitro* 1984; 20(5):376-384.
- (14) Ashby R, Hjortkjaer RK, Stavnsbjerg M et al. SAFETY EVALUATION OF STREPTOMYCES-MURINUS GLUCOSE ISOMERASE. *Toxicology Letters (Shannon)* 1987; 36(1):23-36.
- (15) Bar A, Krul CAM, Jonker D, de VN. Safety evaluation of an alpha-cyclodextrin glycosyltransferase preparation. *Regulatory Toxicology and Pharmacology* 2004; 39(Suppl. 1):S47-S56.
- (16) Bergman A, Broadmeadow A. An overview of the safety evaluation of the *Thermomyces lanuginosus* xylanase enzyme (SP 628) and the *Aspergillus aculeatus* xylanase enzyme (SP 578). *Food Addit Contam* 1997; 14(4):389-398.
- (17) Biziulevichius GA, Arrestov IG. Safety of lysosubtilin per os in mice, rabbits and calves. *Vet Res* 1997; 28(4):385-395.
- (18) Brinch DS, Pedersen PB. Toxicological studies on Laccase from *Myceliophthora thermophila* expressed in *Aspergillus oryzae*. *Regul Toxicol Pharmacol* 2002; 35(3):296-307.
- (19) Broadmeadow A, Clare C, De Boer AS. An overview of the safety evaluation of the *Rhizomucor miehei* lipase enzyme. *Food Addit Contam* 1994; 11(1):105-119.
- (20) Broadwell AH, Baumann L, Baumann P. The 42- and 51-kilodalton mosquitocidal proteins of *Bacillus sphaericus* 2362: construction of recombinants with enhanced expression and in vivo studies of processing and toxicity. *J Bacteriol* 1990; 172(5):2217-2223.
- (21) Bui Q, Geronian K, Gudi R, Wagner V, Kim D, Cerven D. Safety evaluation of marmanase enzyme, produced by *Bacillus lentus*, intended for use in animal feed. *International Journal of Toxicology* 2004; 23(6):398.
- (22) BSr A, Til HP, Timonen M. Subchronic oral toxicity study with regular and enzymatically depolymerized sodium carboxymethylcellulose in rats. *Food Chem Toxicol* 1995; 33(11):909-917.

(23) Cerven D, DeGeorge G, Bethell D. 28-Day repeated dose oral toxicity of recombinant human apolactoferrin or recombinant human lysozyme in rats. *Regulatory Toxicology and Pharmacology* 2008; 51(2):162-167.

(24) Ciofalo V, Barton N, Kretz K, Baird J, Cook M, Shanahan D. Safety evaluation of a phytase, expressed in *Schizosaccharomyces pombe*, intended for use in animal feed. *Regulatory Toxicology and Pharmacology* 2003; 37(2):286-292.

(25) Coenen TM, Schoenmakers AC, Verhagen H. Safety evaluation of beta-glucanase derived from *Trichoderma reesei*: summary of toxicological data. *Food Chem Toxicol* 1995; 33(10):859-866.

(26) Coenen TM, Aughton P, Verhagen H. Safety evaluation of lipase derived from *Rhizopus oryzae*: summary of toxicological data. *Food Chem Toxicol* 1997; 35(3-4):315-322.

(27) Coenen TM, Aughton P. Safety evaluation of amino peptidase enzyme preparation derived from *Aspergillus niger*. *Food Chem Toxicol* 1998; 36(9-10):781-789.

(28) Coenen TM, Bertens AM, de Hoog SC, Verspeek-Rip CM. Safety evaluation of a lactase enzyme preparation derived from *Kluyveromyces lactis*. *Food Chem Toxicol* 2000; 38(8):671-677.

(29) Cook MW, Thygesen HV. Safety evaluation of a hexose oxidase expressed in *Hansenula polymorpha*. *Food Chem Toxicol* 2003; 41(4):523-529.

(30) Deboer AS, Marshall R, Broadmeadow A, Hazelden K. Toxicological Evaluation of Acetolactate Decarboxylase. *J Food Prot* 1993; 56(6):510-517.

(31) Durden DL, Distasio JA. CHARACTERIZATION OF THE EFFECTS OF ASPARAGINASE FROM *ESCHERICHIA-COLI* AND A GLUTAMINASE-FREE ASPARAGINASE FROM *VIBRIO-SUCCINOGENES* ON SPECIFIC CELL MEDIATED CYTO TOXICITY. *International Journal of Cancer* 1981; 27(1):59-66.

(32) Elvig SG, Pedersen PB. Safety evaluation of a glucanase preparation intended for use in food including a subchronic study in rats and mutagenicity studies. *Regulatory Toxicology and Pharmacology* 2003; 37(1):11-19.

(33) Gao C, Zhang A, Lin Y, Han S, Wang L. Relationship between the domain structures of several nuclear receptors and the effect differences of environmental endocrine disrupting chemicals. *Asian Journal of Ecotoxicology* 2007; 2(4):363-374.

(34) Gao F, Jiang Y, Zhou GH, Han ZK. The effects of xylanase supplementation on growth, digestion, circulating hormone and metabolite levels, immunity and gut microflora in cockerels fed on wheat-based diets. *Br Poult Sci* 2007; 48(4):480-488.

(35) Greenough RJ, Everett DJ, Stavnsbjerg M. Safety evaluation of alkaline cellulase. *Food Chem Toxicol* 1991; 29(11):781-785.

(36) Harbak L, Thygesen HV. Safety evaluation of a xylanase expressed in *Bacillus subtilis*. *Food Chem Toxicol* 2002; 40(1):1-8.

(37) Harper AF, Skaggs JH, Veit HP, Kornegay ET. Efficacy and safety of Novo SP938 microbial phytase supplementation of a corn-soybean meal diet fed to growing pigs. *Journal of Animal Science* 1999; 77(SUPPL. 1):174-175.

(38) Hjortkjaer RK, Bille-Hansen V, Hazelden KP et al. Safety evaluation of Celluclast, an acid cellulase derived from *Trichoderma reesei*. *Food Chem Toxicol* 1986; 24(1):55-63.

(39) Hjortkjaer RK, Stavnsbjerg M, Pedersen PB et al. Safety evaluation of esperase. *Food Chem Toxicol* 1993; 31(12):999-1011.

(40) Holcenberg JS, Borella LD, Camitta BM, Ring BJ. HUMAN PHARMACOLOGY AND TOXICOLOGY OF SUCCINYLATED ACINETOBACTER GLUTAMINASE ASPARAGINASE. *Cancer Research* 1979; 39(8):3145-3151.

(41) Hytonen M, Vanhanen M, Keskinen H, Tuoni T, Tupasela O, Nordman H. Pharyngeal edema caused by occupational exposure to cellulase enzyme. *ALLERGY EUR J ALLERGY CLIN IMMUNOL* 1994; 49(9):782-784.

(42)Janer G, Hakkert BC, Piersma AH, Vermeire T, Slob W. A retrospective analysis of the added value of the rat two-generation reproductive toxicity study versus the rat subchronic toxicity study. *Reproductive Toxicology* 2007; 24(1, Sp. Iss. SI):103-113.

(43)Klinge L, Straub V, Neudorf U, Volt T. Enzyme replacement therapy in classical infantile Pompe disease: Results of a ten-month follow-up study. *Neuropediatrics* 2005; 36(1):6-11.

(44)Klinge L, Straub V, Neudorf U et al. Safety and efficacy of recombinant acid alpha-glucosidase (rhGAA) in patients with classical infantile Pompe disease: results of a phase II clinical trial. *Neuromuscul Disord* 2005; 15(1):24-31.

(45)Kondo M, Ogawa T, Matsubara Y, Mizutani A, Murata S, Kitagawa M. Safety evaluation of lipase G from *Penicillium camembertii*. *Food Chem Toxicol* 1994; 32(8):685-696.

(46)Kopetzki E, Lehnert K, Buckel P. Enzymes in diagnostics: Achievements and possibilities of recombinant DNA technology. *Clinical Chemistry* 1994; 40(5):688-704.

(47)Kornegay ET, Skaggs JH, Denbow DM, Larsen CT, Veit HP. Efficacy and safety of Novo SP938 microbial phytase supplementation of a low-P corn-soybean meal diet fed to turkeys. *Poultry Science* 1999; 78(SUPPL. 1):15.

(48)Laake K. ENZYMIC DRUGS. *Side Effects of Drugs Annual* 1980;222-225.

(49)Landry TD, Chew L, Davis JW et al. Safety evaluation of an alpha-amylase enzyme preparation derived from the archaeal order Thermococcales as expressed in *Pseudomonas fluorescens* biovar I. *Regul Toxicol Pharmacol* 2003; 37(1):149-168.

(50)Lane RW, Yamakoshi J, Kikuchi M, Mizusawa K, Henderson L, Smith M. Safety evaluation of tannase enzyme preparation derived from *Aspergillus oryzae*. *Food Chem Toxicol* 1997; 35(2):207-212.

(51)MacKenzie KM, Petsel SR, Weltman RH, Zeman NW. Subchronic toxicity studies in dogs and in utero rats fed diets containing *Bacillus stearothermophilus* alpha-amylase from a natural or recombinant DNA host. *Food Chem Toxicol* 1989; 27(9):599-606.

(52)Modderman JP, Foley HH. Safety evaluation of pullulanase enzyme preparation derived from *Bacillus licheniformis* containing the pullulanase gene from *Bacillus deramificans*. *Regulatory Toxicology and Pharmacology* 1995; 21(3):375-381.

(53)Ohshita K, Nakajima Y, Yamakoshi J, Kataoka S, Kikuchi M, Pariza MW. Safety evaluation of yeast glutaminase. *Food and Chemical Toxicology* 2000; 38(8):661-670.